

Product Name : DynaMarker DIG Labeled Blue Color Marker for Small RNA
(Previous name: DynaMarker Colored DIG Marker for Small RNA)

Code No. : DM270

Range : 20 - 100 bases

Size : 125 μ l (5 μ l \times 25 loadings)

Storage : store at -20 °C

Description :

The DynaMarker DIG Labeled Blue Color Marker for Small RNA consists of colored (Blue) and DIG labeled six single-strand nucleic acids, the apparent molecular weights of which are 20, 30, 40, 50, 75 and 100 bases of RNAs. This marker is suitable for monitoring denaturing polyacrylamide gel electrophoresis and for immunodetection with anti-DIG antibody.

The apparent sizes of bands in DynaMarker DIG Labeled Blue Color Marker for Small RNA are in excellent agreement with sizes of non-stained RNAs, 20, 30, 40, 50, 75 and 100 bases in length (about 95 % accuracy, see Table 1). The DynaMarker DIG Labeled Blue Color Marker for Small RNA is supplied in a ready-to-use mixture and doesn't require heating or addition of a denaturing agent before use.

Storage buffer :

2 mM Tris-HCl (pH8.0), 8mM EDTA, 78 % Formamide

Quality Control :

After 24-hrs incubation of the DynaMarker DIG Labeled Blue Color Marker for Small RNA at 37 °C, no visible degradation of the marker is observed in 15 % polyacrylamide - 7.5 M urea gel electrophoresis.

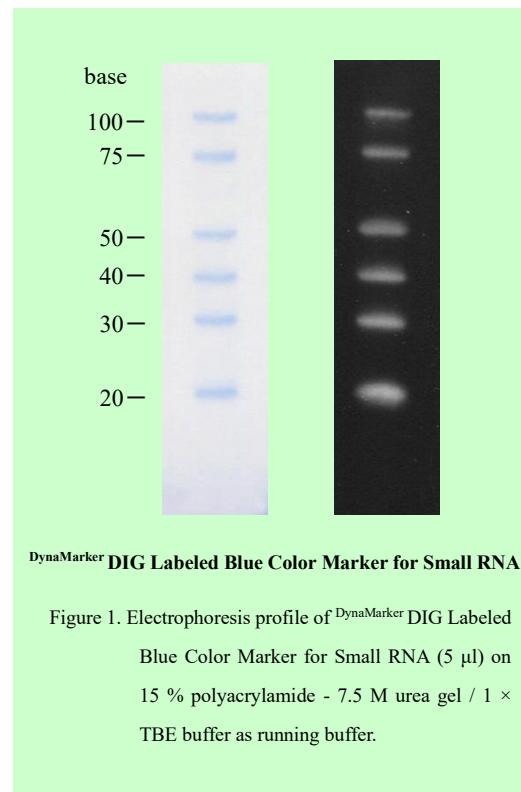
Recommended loading volumes : 5 - 10 μ l

Electrophoresis condition :

Be sure to use this marker on 10 - 15% acrylamide - Urea /1 \times TBE gel and 1 \times TBE as running buffer. Under other conditions, the bands cannot be separated correctly.

Note :

For accurate electrophoretic determination of molecular weights, the DynaMarker Small RNA II (code # DM192) or DynaMarker Small RNA II Easy Load (code # DM197) should be used.



		acrylamide concentration	
		10 %	15 %
DynaMarker Small RNA II + 75 base RNA	100 base	103.9 %	101.3
	75*	104.8	100.6
	50	102.5	101.6
	40	103.0	100.5
	30	99.6	104.5
	20	100.0	102.8

Table 1. This shows apparent molecular weights compared with the DynaMarker Small RNA II, and suitable acrylamide concentrations for electrophoresis of the DynaMarker DIG Labeled Blue Color Marker for Small RNA.

(* 75 base RNA is from a newly synthesized RNA. A 75 base RNA is not included in DynaMarker Small RNA II.)

Sensitivity :

This marker is suitable for chemiluminescent (e.g. CDP-star^{*1}) immunoassay. The sensitivity depends on the length of exposure to high speed instant film (e.g. FP-3000B^{*2}), X-ray film or imaging instrument.

The following figure shows an example.

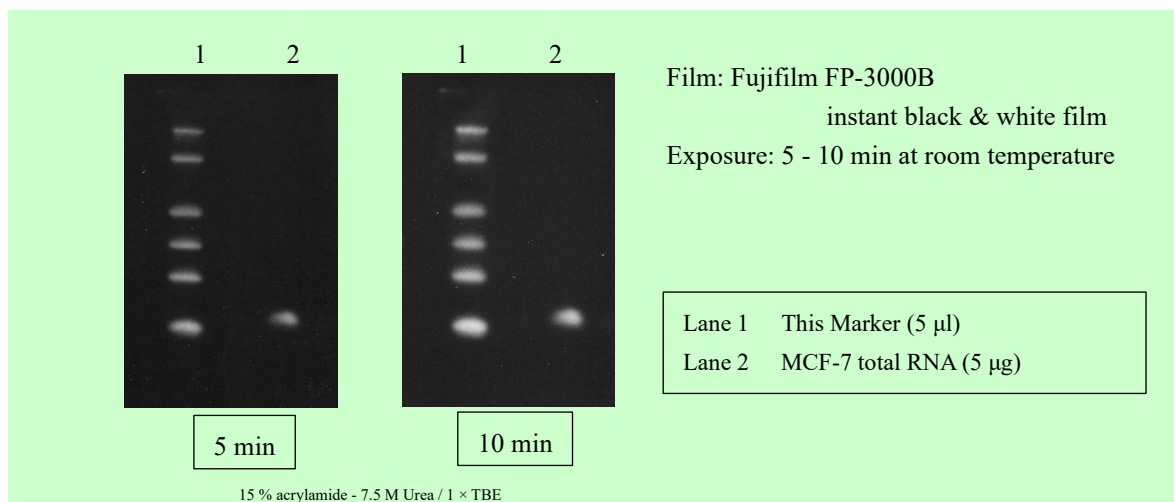


Figure 2. Detection of DynaMarker DIG Labeled Blue Color Marker for Small RNA and hsa-miR-21.

5 µl of DynaMarker DIG Labeled Blue Color Marker for Small RNA and 5 µg of MCF-7 total RNA were blotted onto nylon membrane, and the hsa-miR-21 was hybridized with the DIG labeled DNA probe (2.5 nM). This marker and the DIG labeled DNA probe were detected with anti-DIG-AP antibody and CDP-star.

*1: CDP-star is a trademark of Tropix, Inc.

*2: FP-3000B is a product of Fujifilm Corp.

Recommended usage :

The ^{DynaMarker} DIG Labeled Blue Color Marker for Small RNA is suitable for monitoring denaturing acrylamide gel electrophoresis and blotting onto membrane. One example is shown below:

• Electrophoresis and blotting of ^{DynaMarker} DIG Labeled Blue Color Marker for Small RNA

1) Preparation of 12.5 % polyacrylamide - 7.5 M urea gel

40 % acrylamide : bis solution	6.25 ml
Urea	9.0 g
10 × TBE	2.0 ml
H ₂ O	to 20 ml

After urea is dissolved completely, add 20 µl of TEMED and 100 µl of 10 % ammonium persulfate. Mix quickly then pour the gel into the mold of a vertical gel apparatus.

2) Loading and electrophoresis.

Thaw the ^{DynaMarker} DIG Labeled Blue Color Marker for Small RNA completely before use. Load the denatured RNA sample and 5 µl of ^{DynaMarker} DIG Labeled Blue Color Marker for Small RNA into a well and run the gel using 1 × TBE electrophoresis buffer at 200 V.

3) Transfer the ^{DynaMarker} DIG Labeled Blue Color Marker for Small RNA and RNA from gel to membrane (Figure 3).

3-1) Cut a piece of positive charged nylon membrane slightly larger than the gel. Soak the membrane and four sheets of blotting paper of appropriate size in 1 × TBE buffer.

3-2) Place two sheets of blotting paper on the anode platform of the transfer cell.

3-3) Place the membrane on top of the blotting paper.

3-4) Transfer the gel from the glass plate to the top of the membrane and press out any air bubbles.

(*Make sure that there are no air bubbles between the membrane and the gel.)

3-5) Place another two sheets of blotting paper onto the gel and set the cathode assembly.

3-6) Transfer for 30 - 60 min at 2 mA/cm².

3-7) After ensuring the marker has transferred successfully onto the membrane, remove both paper and gel. Rinse the membrane in 2 × SSC.

3-8) Fix the RNA to the membrane with a UV crosslinker.

3-9) Carry out northern hybridization (Figure 4).

References:

- Joseph Sambrook, and David W. Russell (2001) Molecular Cloning: A Laboratory Manual, 3rd ed., Cold Spring Harbor Laboratory Press.
- Frederick M. Ausubel, Roger Brent, Robert E. Kingston, David D. Moore, J. G. Seidman, John A. Smith, and Kevin Struhl (1994—) Current Protocols in Molecular Biology, John Wiley & Sons, Inc.
- Sang Woo Kim, Zhihua Li, Patrick S. Moore, A. Paula Monaghan, Yuan Chang, Mark Nichols and Bino John (2010) A sensitive non-radioactive northern blot method to detect small RNAs. Nucleic Acids Research. **38**(7): e98

